

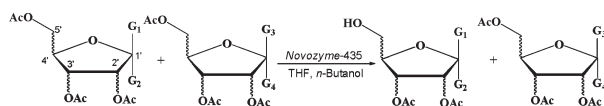
## Biocatalytic Separation of *N*-7/*N*-9 Guanine Nucleosides

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**$\beta$ -D-ribofuranosyl guanine:**  
G<sub>1</sub> = *N*<sup>2</sup>-isobutanoylguanin-9-yl, G<sub>2</sub> = H, C-2' & C-3' OAc ( $\alpha$ ), C-4' CH<sub>2</sub>OAc ( $\beta$ )  
G<sub>3</sub> = *N*<sup>2</sup>-isobutanoyl-7-yl, G<sub>4</sub> = H, C-2' & C-3' OAc ( $\alpha$ ), C-4' CH<sub>2</sub>OAc ( $\beta$ )  
 **$\alpha$ -D-arabinofuranosyl guanine:**  
G<sub>1</sub> = H, G<sub>2</sub> = *N*<sup>2</sup>-isobutanoylguanin-9-yl, C-2' OAc ( $\beta$ ), C-3' OAc ( $\alpha$ ), C-4' CH<sub>2</sub>OAc ( $\beta$ )  
G<sub>3</sub> = H, G<sub>4</sub> = *N*<sup>2</sup>-isobutanoylguanin-7-yl, C-2' OAc ( $\beta$ ), C-3' OAc ( $\alpha$ ), C-4' CH<sub>2</sub>OAc ( $\beta$ )  
 **$\alpha$ -L-arabinofuranosyl guanine:**  
G<sub>1</sub> = *N*<sup>2</sup>-isobutanoylguanin-9-yl, G<sub>2</sub> = H, C-2' OAc ( $\alpha$ ), C-3' OAc ( $\beta$ ), C-4' CH<sub>2</sub>OAc ( $\alpha$ )  
G<sub>3</sub> = *N*<sup>2</sup>-isobutanoylguanin-7-yl, G<sub>4</sub> = H, C-2' OAc ( $\alpha$ ), C-3' OAc ( $\beta$ ), C-4' CH<sub>2</sub>OAc ( $\alpha$ )

Vorbrüggen coupling of trimethylsilylated 2-*N*-isobutanoylguanine with peracetylated pentofuranose derivatives generally gives inseparable *N*-7/*N*-9 glycosyl mixtures. We have shown that the *two* isomers can be separated biocatalytically by Novozyme-435-mediated selective deacetylation of the 5'-*O*-acetyl group of peracetylated *N*-9 guanine nucleosides.

The discovery of 9-[(2-hydroxyethoxy)methyl]guanine (acyclovir) against herpes simplex type 1 and type 2 viruses and its low mammalian toxicity triggered the synthesis of a series of guanine nucleosides, e.g., penciclovir, famciclovir, valaciclovir, valganciclovir, abacavir, etc., for the treatment of various viral diseases.<sup>1,2</sup> The most problematic chemistry and difficulties in manipulation of all five common bases found in DNA and RNA occur with the polyfunctional guanine ( $pK_{a1} - 1.7$ ,  $pK_{a2} - 9.2$ ) nucleosides and nucleotides. The coupling of guanine with peracetylated sugar derivatives generally produces *N*-7/*N*-9 isomeric mixtures of nucleosides

(1) Elion, G. B.; Furman, P. A.; Fyfe, J. A.; Miranda, P. De.; Beauchamp, L.; Schaeffer, H. J. *Proc. Natl. Acad. Sci. U.S.A.* **1977**, *74*, 5716–5720.

(2) (a) Herdewijn, P. Modified Nucleosides. In *Biochemistry, Biotechnology and Medicine*; Wiley-VCH: New York, 2008. (b) Chu, C. K. *Antiviral Nucleosides: Chiral Synthesis and Chemotherapy*; Elsevier: New York, 2003.

that are difficult to separate.<sup>3,4</sup> The changes in experimental variables and the use of a selectively modified guanine moiety, such as 2-*N*-acetyl-6-*O*-diphenylcarbonylguanine in nucleoside coupling reactions, affect the isomeric ratio but do not eliminate the formation of the *N*-7 isomer together with the desired *N*-9 isomer.<sup>5</sup> In this paper, we report the synthesis of guanine nucleosides (mixture of 9- and 7-glycosyl derivatives) derived from D-ribose, D-arabinose, and L-arabinose sugars and for the first time their highly efficient separation mediated by Novozyme-435 lipase-catalyzed removal of one of the acetoxy functions of the peracetylated *N*-9 guanine nucleosides.

The coupling of 2-*N*-isobutanoylguanine (**2**)<sup>6</sup> with 1,2,3,5-tetra-*O*-acetyl-D-ribofuranose (**3**),<sup>7,8</sup> 1,2,3,5-tetra-*O*-acetyl-D-arabinofuranose (**4**),<sup>9</sup> or 1,2,3,5-tetra-*O*-acetyl-L-arabinofuranose (**5**)<sup>8–10</sup> in the presence of TMSOTf as Lewis acid catalyst following a standard Vorbrüggen<sup>5,11</sup> coupling protocol afforded mixtures of 2,3,5-tri-*O*-acetylated 9- and 7- $\beta$ -D-ribofuranosylguanines **6** and **7**, 9- and 7- $\alpha$ -D-arabinofuranosylguanines **8** and **9**, and 9- and 7- $\alpha$ -L-arabinofuranosylguanines **10** and **11** in ratios of 87:13, 63:37, and 76:24, respectively, in 60–65% yields (Scheme 1 and Table 1). The ratio of regioisomers *N*-9 and *N*-7 in the above guanine nucleoside mixtures **6** and **7**, **8** and **9**, and **10** and **11** were calculated on the basis of the integration of the corresponding anomeric protons in the <sup>1</sup>H NMR spectra (400 MHz) of the mixtures (Table 1). Our various attempts of separation of *N*-9 and *N*-7 guanine nucleosides from the mixtures **6** and **7**, **8** and **9**, and **10** and **11** by repeated column chromatography on silica gel were unsuccessful.

Some lipases have been found to selectively acylate/deacylate primary hydroxyl over secondary hydroxyl group(s) of sugars<sup>12</sup>

(3) Zhong, M.; Robins, M. J. *Tetrahedron Lett.* **2003**, *44*, 9327–9930.

(4) (a) Schaffer, H. J.; Beauchamp, L.; Miranda, P. de.; Elion, G. B.; Bauer, D. J.; Collins, P. *Nature* **1978**, *272*, 583–585. (b) Robins, M. J.; Hatfield, P. W. *Can. J. Chem.* **1982**, *60*, 547–553. (c) Boryski, J.; Golankiewicz, B. *Nucleosides Nucleotides* **1987**, *6*, 385–386. (d) Matsumoto, H.; Kaneko, C.; Yamada, K.; Takeuchi, T.; Mori, T.; Mizuno, Y. *Chem. Pharm. Bull.* **1988**, *36*, 1153–1157. (e) Boryski, J. *J. Chem. Soc., Perkin Trans. 2* **1997**, 649–652. (f) Clair, A. S.; Xiang, G.; McLaughlin, L. W. *Nucleosides Nucleotides* **1998**, *17*, 925–937. (g) Boryski, J.; Golankiewicz, B. *Nucleosides Nucleotides* **1989**, *8*, 529–536. (h) Geen, G.; Grinter, T. J.; Kinsey, P. M.; Jarvest, R. L. *Tetrahedron* **1990**, *46*, 6903–6914. (i) Izawa, K.; Shiragami, H. *Pure Appl. Chem.* **1998**, *70*, 313–318. (j) Li, N. -S.; Piccirilli, J. A. *Synthesis* **2005**, *17*, 2685–2870. (k) Ferenc, G.; Kele, Z.; Kovács, L. *Rapid Commun. Mass Spectrom.* **2005**, *19*, 236–240.

(5) (a) Vorbrüggen, H.; Krolkiewicz, K.; Bennua, B. *Chem. Ber.* **1981**, *114*, 1234–1255. (b) Robins, M. J.; Zou, R.; Guo, Z.; Wnuk, S. F. *J. Org. Chem.* **1996**, *61*, 9207–9212. (c) Garner, P.; Ramakanth, S. *J. Org. Chem.* **1988**, *53*, 1294–1298. (d) Zou, R.; Robins, M. J. *Can. J. Chem.* **1987**, *65*, 1436–1437. (e) Cheung A. W.-H.; Sidduri, A.; Garofalo, L. M.; Goodnow, R. A., Jr. *Tetrahedron Lett.* **2000**, *41*, 3303–3307.

(6) Milecki, J.; Foldesi, A.; Fischer, A.; Adamiak, R. W.; Chattopadhyaya, J. *J. Labelled Compd. Radiopharm.* **2001**, *44*, 763–783.

(7) Wright, G. E.; Dudycz, L. W. *J. Med. Chem.* **1984**, *27*, 175–181.

(8) Shi, C.-J.; Zhang, J.; Fu, J.; Tang, J. *Lett. Org. Chem.* **2006**, *3*, 932–935.

(9) Gupta, P.; Maity, J.; Shakya, G.; Prasad, A. K.; Parmar, V. S.; Wengel, J. *Org. Biomol. Chem.* **2009**, *7*, 2389–2401.

(10) Kam, B. L.; Barascut, J. L.; Imbach, J. L. *Carbohydr. Res.* **1979**, *69*, 135–142.

(11) Vorbrüggen, H.; Höfle, G. *Chem. Ber.* **1981**, *114*, 1256–1268.

(12) (a) Prasad, A. K.; Kalra, N.; Yadav, Y.; Kumar, R.; Sharma, S. K.; Patkar, S.; Lange, L.; Wengel, J.; Parmar, V. S. *Chem. Commun.* **2007**, 2616–2617. (b) Maity, J.; Shakya, G.; Singh, S. K.; Ravikumar, V. T.; Parmar, V. S.; Prasad, A. K. *J. Org. Chem.* **2008**, *73*, 5629–5632. (c) Sharma, R. K.; Aggarwal, N.; Arya, A.; Olsen, C. E.; Parmar, V. S.; Prasad, A. K. *Indian J. Chem.* **2009**, *48B*, 1727–1731.